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UNIT-IV

Plant Tissue Culture

OR

In vitro culture

OR

Sterile culture or Aseptic

Definition

Plant tissue culture is a method or technique used to isolate parts (protoplasts, cells, tissue & organ) and grow them on a nutritive medium under aseptic condition in a controlled space so that they can grow and develop into complete plant.

OR

Plant tissue culture is a collection of technique used to maintain or growth of plant cell tissue or organ on a ^{nutritive} nutrient medium of 1/4n composition under sterile condition in

OR

Tissue is in vitro cultivation of plant cell or tissue under aseptic condition and controlled environment condition in liquid or semisolid well defined culture medium for the production of 1st & 2nd metabolites OR regenerate a new plant.

Advantages-

- Availability of raw material
- Quickly produces mature plant
- reproduction of disease free plants
- crop improvement by somatic hybridization or / production of hybrid.
- The regeneration of whole plants from plant ^{cell} ~~stems~~ that have been modified
- Immobilization of cells
- Easy purification of the compounds
- Tissue culture can be used for tracing the biosynthetic pathway of secondary metabolites by using labeled precursors in the culture medium
- Patent rights

Historical Development of Plant Tissue Culture

Year/About	Name/Author	Results
1902	Haberlandt (Father of Plant Tissue Culture)	1 st cultivation experiment with isolated plant cell, cell growth but no cell division obtained
1904	Hanning	Established of the embro. culture of the first time
1922	Kotte & Rabbiris	In vitro cultivation of Root tips, No permanent culture obtained
1934	White	successful culture of Tomato roots in a long time he replaced the yeast extract in a medium contain inorganic salt and sucrose with 3 vitamines

(4)

Explant

1934

Gravmet

First permanent callus culture using Vitamine B and Oxgin

1946

Ball

Microw-propagation, Development of stem tips and branches

1941

Van overbeek

The use of condensed water for the young embryo culture in datura plant

1954

Muir et al

First suspension culture of Singal cell or callus

1955

Mothur & Kala

First reports of Secondary metabolites production in liquid medium

1957

Stoog & Miller

Proposed the concept of hormonal control of original production

1958	Mahekwari & Rangaswamy	Regeneration of somatic embryo of circhus ovules
1960	Bergmann	cell colonies (exact copy) obtained from <u>Birgal</u> culture cell plated in an agar medium
1960	Reinert Steward	growth and development of carrot suspension culture, enzymatic degradation of cell wall via somatic embryogenesis
1967	Cruha & Maheswari	The discovery of first Hypoid plant by immature polygamit in <u>Datura plant</u>
1970	Power et al	Demonstrated <u>intra and interspecific fusion</u> between the <u>potoplast of different plant roots</u>
1978	Mejcher's et al	first <u>intergeneric hybridization</u> between <u>tomato and potato plant</u>

Basic requirement for a tissue culture of laboratory -

- For the successful achievement of any type of tissue culture laboratory should have the following basic facilities -

①. Equipment & Apparatus -

- Main different kinds of vessels may be used for growing culture.
For the successful growth of any tissue it requires wide mouth conical flask (Erlenmeyer flask). In addition to the culture vessels, glasswares such as graduated pipellets, measuring cylinder, Beaker, funnel & petri dishes are all so required for making preparation.
- A spirit burner or gas micro burner for flame sterilization are also used.
- Autoclave for sterilizing culture media, Hot air oven for sterilizing glasswares are also used.
- Scissors & forceps are also used for the preparation of explant.
- An equivation chamber, or laminar air flow with U.V. fitted light are also used for aseptic transfers of explant into culture medium.

② Washing & storage facilities -

A good PTC laboratory should have fresh water supply & disposal of the waste water.

③ Media preparation Room -

Media preparation room should have sufficient space to accommodate chemical, labwares, chemical vessels & equipment required for weighing & mixing, hot plate, pH meter, water bath, micro wave oven, autoclave & domestic pressure cooker.

④ Aseptic Chamber -

- For the transfer of culture into sterilized media, contaminant environment is mandatory.
- Now a days modern laboratory have laminar air flow cabinet having vertical or horizontal air flow.
- The air coming out of the fine filter (0.3µm HEPA filter) is ultra clean & having adequate velocity to prevent micro-contamination of the working area.

⑤ Incubation Room -

Air conditioner or air room heater required to the maintain temp at

25°C

important

- Light is adjusted in the term of photo-period duration.

① Culture media - Nutrient media -

Nutrition requirement for optimal growth of a tissue culture may vary with the species. Even tissue from different parts of a plant may have different requirement for the favorable growth of plant cell.

② Media Composition -

To maintain the vital function of a culture the basic medium contains inorganic nutrients, organic components, growth regulators & utilizable carbon sources & a gelling agents.

③ Inorganic Nutrient -

Mineral elements play very important role in the growth of a plant.

ex - Mg^{2+} , is a part of chlorophyll.

Ca^{2+} , is a component of cell wall.

N, is an important element of amino acid.

Iron, Zinc, Molybdenum are parts of

certain enzyme.

- Essentially about 15 elements are required for the favorable growth of a plant tissue in a culture media.

- Elements required in the life of a plant greater than 0.5 m.mol/lit or referred as macro-nutrient ~~ie~~ those less than 0.5 m.mol/lit as micro nutrient.

- The macro-nutrient elements includes 6 major elements, Nitrogen, P, Ca, Mg, S, K

- The micro-nutrients are required in small quantities but essential for proper growth of plant cell or tissue these nutrient are B, Co, Fe, Mn, Zn & Mo.

© • Organic Nutrients -

Most cultured plant cells are capable of synthesizing essential vitamins but not in sufficient amount.

- To achieve best growth it requires essential vitamins such as -
 Vit - B₁ - (Thiamine)
 Vit - B₆ - (Pyridoxine)
 Vit - B₅ - (Pantothenate).

• Carbon sources -

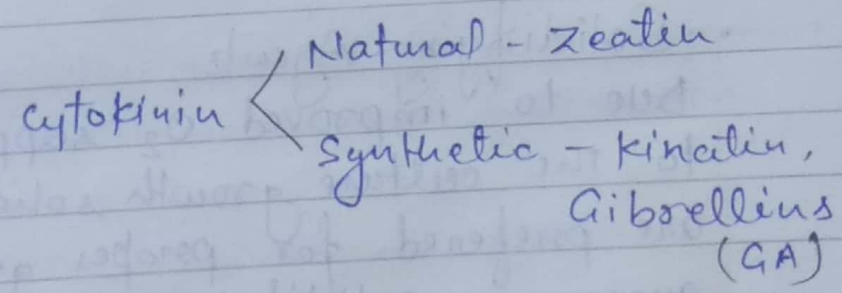
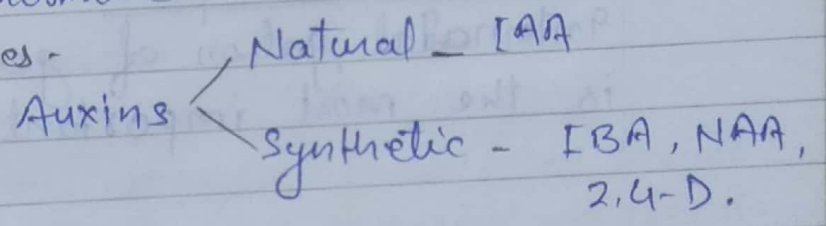
It is essential to supplement the tissue culture media with utilizable source of carbon.

- The most commonly used carbon source is sucrose at concⁿ 2-5%.
- Glucose & fructose also known to be used for growth of some tissues.
- In general dicotyledonous roots grow better with sucrose where as monocots do best with glucose.

(d) Growth Hormones

In addition to the nutrients it is generally necessary to add one or more growth hormone.

Examples -



- Auxin are use elongation of stem and internode and apical dominance and rooting etc.
- In tissue culture auxin induces all division and stimulate root formation
- IAA - Indole-3-acetic acid.
- IBA - Indole-3-butyric acid.
- NAA - Naphthalene acetic acid.
- 2,4-D - 2,4-Dichloro phenoxy acetic acid.

Cytokinin -

It is a phytohormone naturally in plant which are to start cell division.

Cytokinin $\left\{ \begin{array}{l} \text{Natural - Zeatin.} \\ \text{Synthetic - kinetin.} \end{array} \right.$

Gibrellins - (GA)

- GA are GA-3 (Gibberellic acetic acid-3) is the most common form of GA.
- It is used for parthenocarpy and apical dominant in plant.
- Internodal gation of genetically plant is the most important property of GA.

Solidifying Agents

Due to improved O_2 supply & support to the culture growth solid media are preferred for proper growth of cell. common solidifying agents are - Agar, Gelatin, alginate, Carrageenan, are used in culture media.

Types of plant tissue culture -

- ① Root tip culture.
- ② Leaves tip culture.
- ③ Shoot tip culture.

- ④. Anther & pollen grain culture.
- ⑤. Ovary & Embryo culture.
- ⑥. Protoplast culture.
- ⑦. Callus culture.
- ⑧. Suspension culture.

①. Root Tip culture -

Tip of the lateral root of sterilized excised & transfer to fresh medium.

The lateral root continue to grow ~~aerobites~~ provides several roots after few days it becomes a mature plants.

②. Leaves Culture -

Leaves may be detached / cut from shoots, surface sterilized and placed on a semi-solid medium where they will remain in a healthy condition for a long period.

③. Shoot tip culture -

The excised shoot tips of many plant species can be cultured on a simple nutrient medium under sterile condition.

④. Anther and pollen culture -

Young flower buds are removed from the plants and surface sterilize. The anther are carefully excised and transferred to an appropriate nutrient medium.

5) Ovule and Embryo culture -

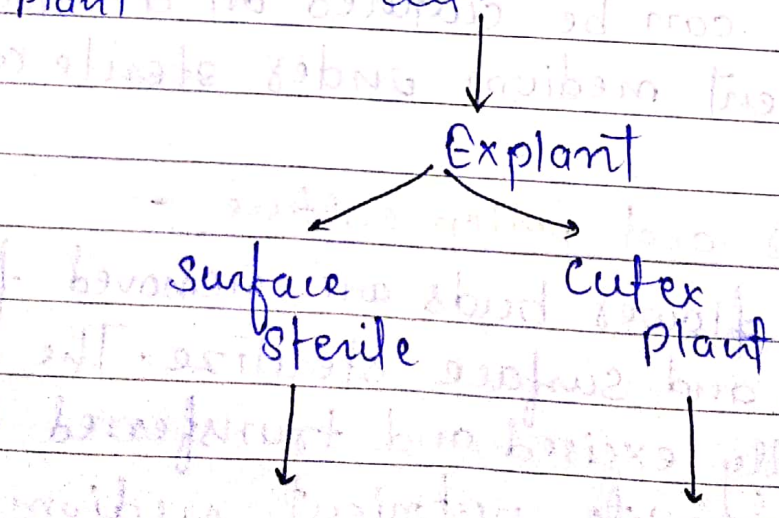
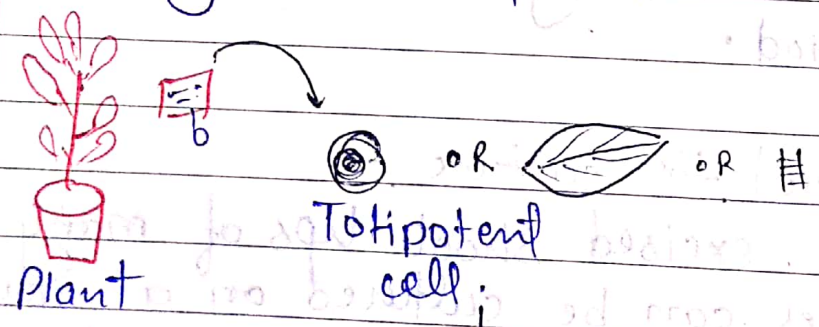
- Embryos are dissected from the ovule and put into the culture medium.
- The seeds are treated with 70% alcohol for about 2 min. washed with sterile distilled water followed by surface sterilizing agent for specific period of time.

~~Exp -~~

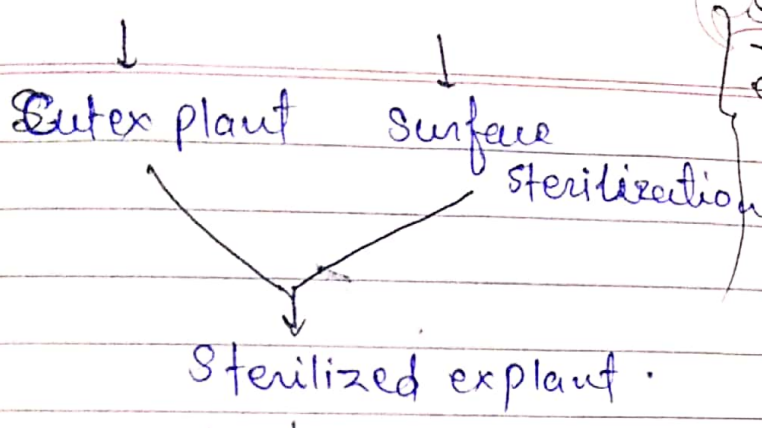
Systemic procedure of Plant tissue Culture/Technique -

Explant - Any part of the plant body which are used to establish a culture.

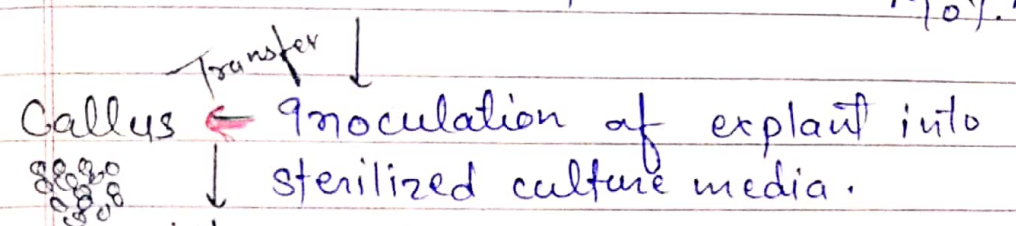
- Explant should be disease free, young & healthy and totipotent.



Date _____
Sterilizing Agent



- eg - Bromine water.
- Benzalkonium chloride.
- H₂O₂
- AgNO₃.
- 70% Alcohol.



into BOD for at least one week.

Incubation (Added phyto hormone if required before incubation.)

Organogenesis.

Rooted plant lets

Hardening

(Acclimatization) grow in soil

Transfer in Polyhouse or green house.

New plant.

Composition of MS Media -
(Musashi S1000 Media)

Macroelement

<u>Ingredients</u>	<u>Quantity (mg / lit)</u>
NH_4NO_3	1650
KNO_3	1900
$\text{CaCl}_2 \cdot 2\text{H}_2\text{O}$	440
$\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$	370
KH_2PO_4	170

Micro Element

<u>Ingredient</u>	<u>Quantity (mg/lit)</u>
KI	0.83
H ₂ BO ₃	6.20
MnSO ₄ · 4H ₂ O	22.30
Na ₂ BO ₃ · 2H ₂ O	0.25
ZnSO ₄ · 4H ₂ O	8.60
CaSO ₄ · 5H ₂ O	0.025
Na ₂ EDTA · 2H ₂ O	37.30
FeSO ₄ · 7H ₂ O	27.80

Amino acid & vitamine

<u>Ingredient</u>	<u>Quantity</u>
Inositol	100
Glycine	2
Thiamine	0.1
Pyridoxin	0.1
Nicotinic Acid	0.5

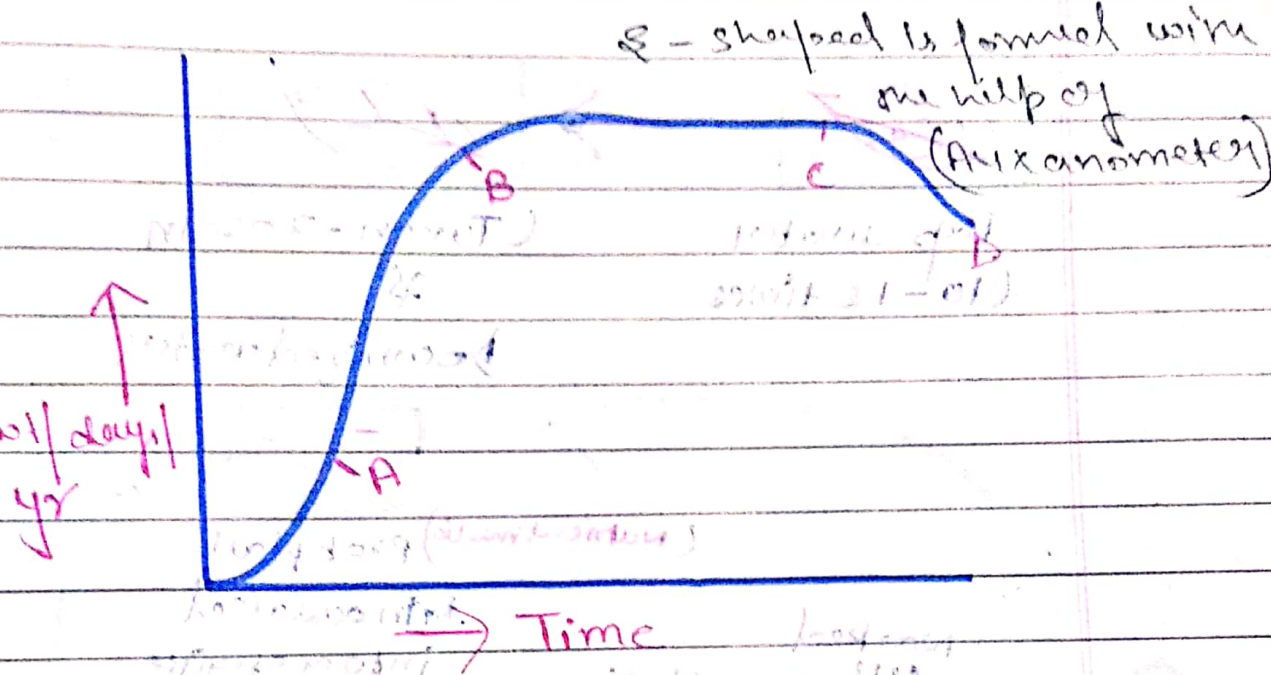
Phytohormones

<u>Ingredient</u>	<u>Quantity</u>
Kinetin	0.4-10
IAA	0.30

Carbon Sources

<u>Ingredient</u>	<u>Quantity</u>
Bovine	30
Agar 201	10.00
1001	5.00
001	0.50
00E	0.50
001	1.00

Growth Profile Measurement



0 - A = Lag Phase

A - B = Log Phase / Exponential

B - C = Steady state phase

C - D = Decline phase or senescence

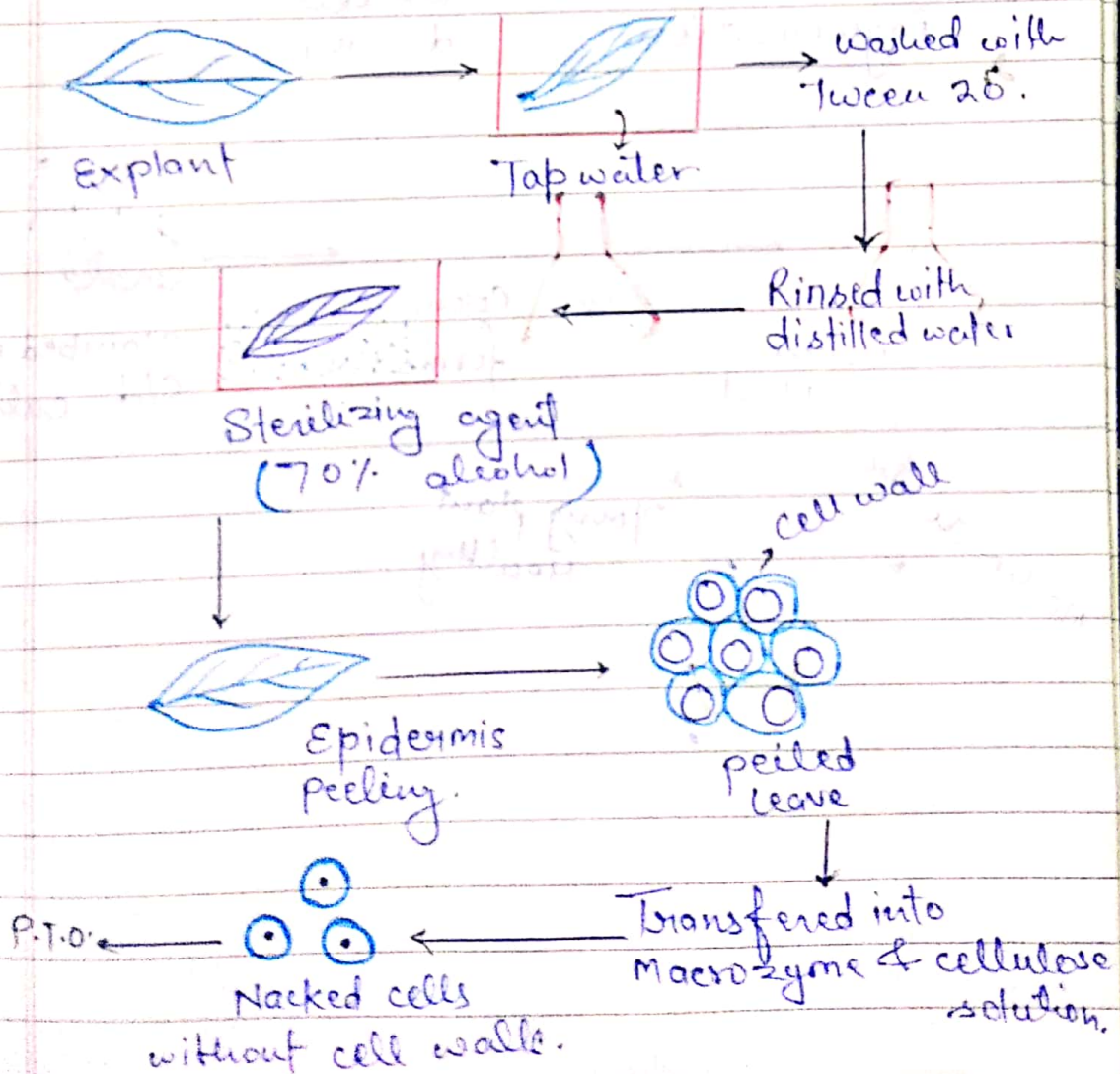
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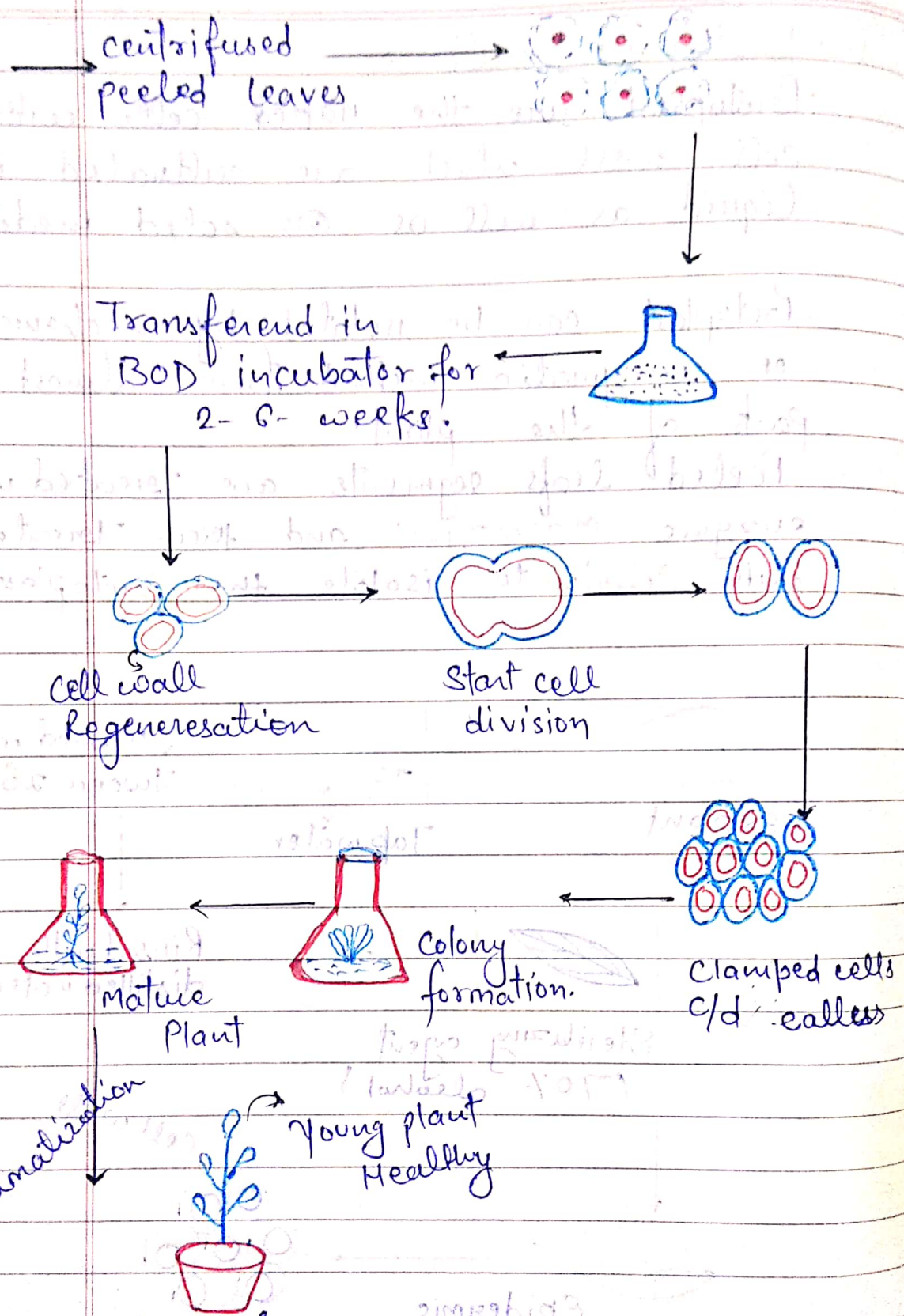
②. Protoplast Culture -

• Protoplast are the naked cells without cell wall which are cultivated in liquid as well as on solid media.

• Protoplast can be isolated by mechanical or enzymatic method from almost all parts of the plant.

Peeled leaf segments are treated with enzyme Macrozyme and then treated with Cellulose to isolate the protoplast.





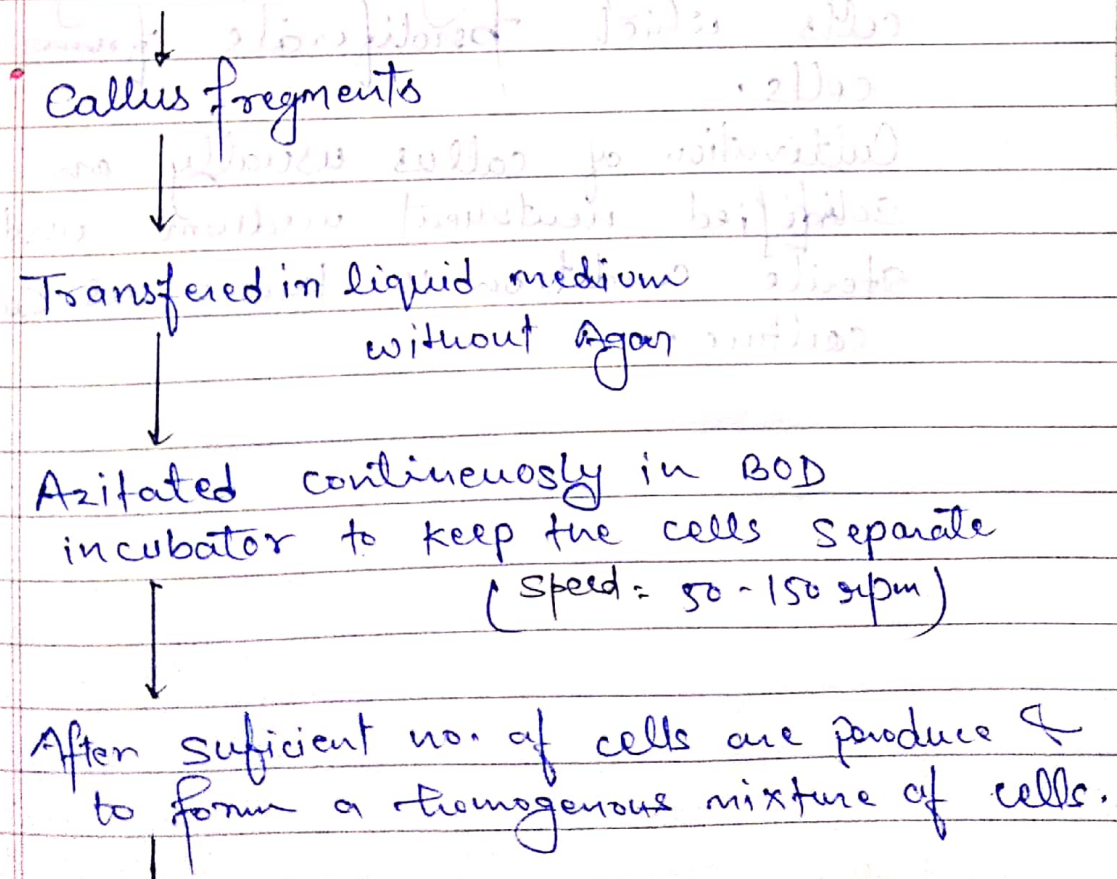
⑦. Suspension Culture -

I. Suspension Culture contains a uniform suspension of separate cells in liquid medium. For the preparation of suspension culture, callus fragments is transferred to liquid medium (without Agar).

which is aerated continuously to keep the cells separate.

The culture maintain medium homogeneous by stirrer speed (50-150 rpm).

After sufficient no. of cells are produced and ~~sub~~-culturing can be done & freshly prepared ~~directly~~ solid medium.



↓
Start Sub-culturing / in freshly prepared solid medium i.e. The single cell is inoculated in fresh medium.

↓
Multiply

↓
Growth of cells.

↓
Organogenous

↓
whole plant.

⑧. Callus Culture -

Callus is an amorphous aggregate of loosely arranged parenchymatous cells which proliferate from mother cells.

Cultivation of callus usually on solidified nutrient medium under sterile condition is known as Callus culture.

Application of plant tissue culture -

Plant tissue culture technique has been used in almost all the field of bioscience.

Major ~~Megase~~ application are the listed below -

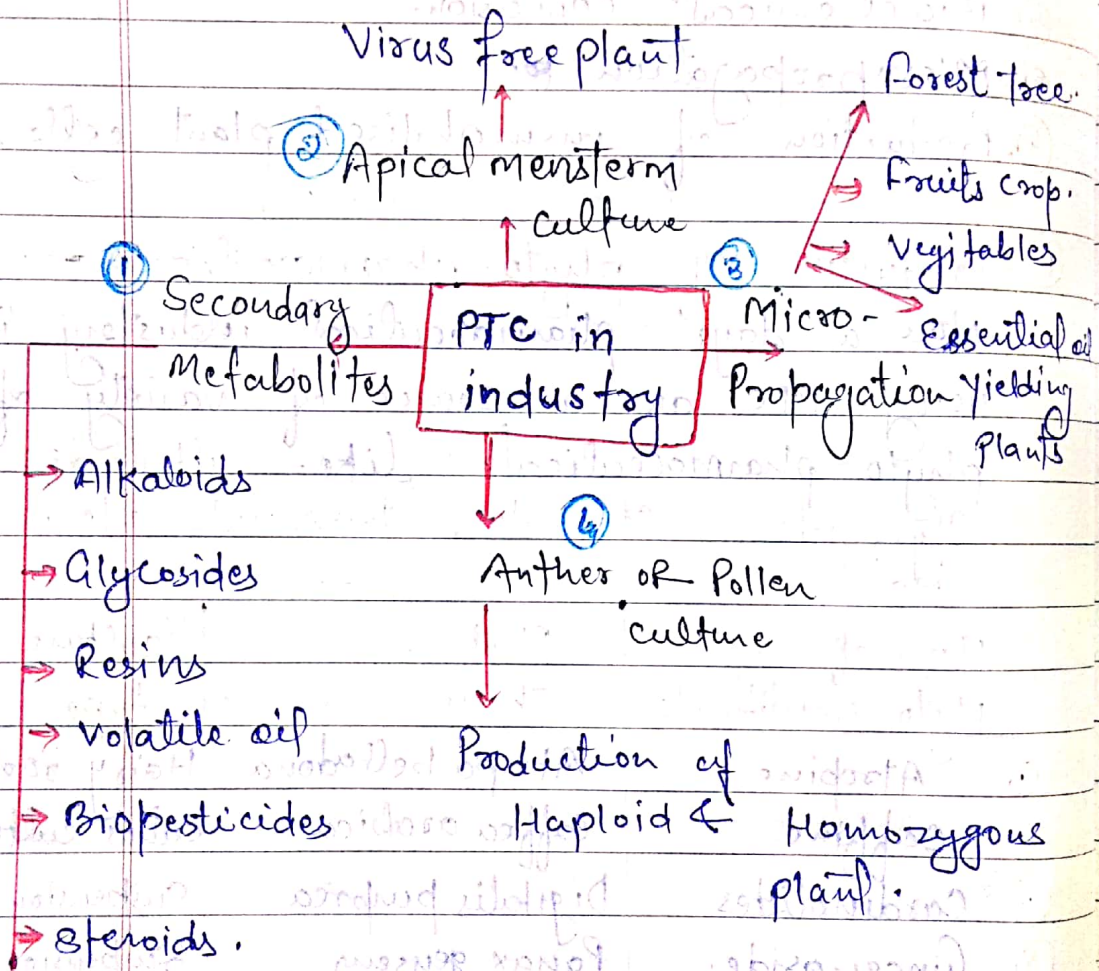
- ①. Production of phyto-pharmaceuticals.
- ②. Biochemical conversion.
- ③. Micropropagation.
- ④. Production of immobilised plant cells.

①. Production of phyto-pharmaceuticals -

Now a days pharmaceutical industry is using PTC as a source of variety of phyto-pharmaceuticals like - alkaloids, glycosides, steroids, terpenoids etc.

Name of Phyto-constituents	Plant Species	Culture type
① Atropine	<i>Atropa belladonna</i>	Hairy root.
② Caffeine	<i>Coffea arabica</i>	callus culture.
③ Cardenolides	<i>Digitalis purpurea</i>	Suspension "
④ Ginsenoside.	<i>Panax ginseng</i>	Suspension "
⑤ Morphine	<i>Papaver Somniferum</i>	Suspension "
⑥ Nicotine	<i>Nicotiana tobacum</i>	Suspension "
⑦ Quinine & ⑧ Quinolone	<i>Cinchona officinalis</i>	callus & Suspension "
⑨ Reserpine	<i>Rauwolfia serpentina</i>	Suspension "

①. Industrial production of 2^o metabolites
Majority of noble medicinal agents are produced by plant tissue culture technique from various type of medicinal plant.



②. Biochemical conversions - or
Biotransformations -

The conversion of small part of a chemical molecule by means of biological system is termed as Biotransformation.

Eg - Podophyllum peltatum in semi continuous culture can produce anticancer drug by biotransformation of synthetic Dibenzyl butanolate to lignan suitable for conversion to etoposide

③. Micropropagation - or

Clonal propagation

Clonal propagation is the technique for rapid production of a large no. of identical clones within a short duration in a limited spaces

④. Immobilization of plant cells -

Immobilization of plant cells or enzymes has increase the utilization of plant cell biotechnology for the production of pharmaceuticals.

The plant cells can be immobilized by using alginates. Polyacrylamide, Polyurethane.

Such methodology may be implemented in case of enzymes which are used as solid support for plant cells.

6

Other application of plant tissue culture

(25)

- 1) Production of genetically variable plant
- 2) Virus eradication (apical meristem are generally virus free or very low conc. virus)
- 3) The apical meristem is the only way to obtain a clone of various ^{disease} free plant
- 4) Study of crown gall by plant tissue culture
- 5) Importance of tissue culture in biotechnology
- 6) production of transgenic plant
- 7) Production genetically variable plants

(26)

Page No. _____
Date: _____

Factor Influence In PTC

→ Size of explant

→ PTC is generally depended upon the size of explant the large explant is consist parenchyma have greater regenerative than small explant

→ Source of explant

The source of explant crucial in important in determining the potential of organogenesis explant may taken any part of plant shoot, stem and leaves. Meristematic tissue combom

→ Age of explant

The physiological age of explant is another factor which often play an important role in organogenesis. The young tissue are more suitable than mature tissue

→ Seasonal variation

→ Quality and Intensity of light

The quality and intensity of light incidence and source may play an effective role in the promotion of organogenesis

Normally plant tissue culture take place in illumination about 2000 to 3000 lux for photo period of 16 hr day and 8 hr night.

→ Temperature

Most tissue culture are grown successfully and temperature around 25°C in a no. of bulbs parts species, the optimum temp may be lower than 15°C

→ Phytohormones

When the conc. of cytokin are high relative to auxin, shoot are induced. when the conc. of cytokin are low relative to auxin roots are induced.

→ Culture Medium

The essential compounds of plant tissue culture is the macrosalt along with vitamins amino acid carbohydrate are also required for growth and development of plant cell with high conc. agar the nutrient medium become more thick does not allow the diffusion of the growing tissue so the conc. of agar also play a critical role in organogenesis



→ pH of the Medium.

The pH of culture medium is generally adjusted b/w 5.6 to 5.7 before sterilization. The pH is another factor that may have a determining role in organogenesis.

→ Age of Culture Medium

The young and freshly prepared culture medium is generally well for the faster & favourable growth of cell or tissue.

→ In the case of primary culture, the medium should be changed frequently to avoid the accumulation of waste products and to provide fresh nutrients.

→ The pH of the medium should be maintained at a constant level throughout the culture period.

→ The concentration of the medium should be adjusted according to the requirements of the cells or tissues being cultured.